ANA HUMAN REFERENCE SERUM #3

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ARTHRITIS FOUNDATION - CENTERS FOR DISEASE CONTROL REFERENCE SERUM FOR
FLUORESCENT ANTINUCLEAR ANTIBODY (SPECKLED PATTERN)

Intended Use

For in vitro immunodiagnostic use as a reference human serum giving a speckled pattern in the fluorescent antinuclear antibody (FANA) test. The speckled pattern obtained is due to antibodies to U1-RNP, SSB/La and SSA/Ro that are present in this serum.

Description of Reference Serum

Citrated plasma from a single donor was made 0.01 M CaCl₂ and 0.013 M epsilon-amino-caproic acid, allowed to clot at 4°C overnight, centrifuged, and frozen at -70°C until lyophilization. Volumes of 0.50 ml were dispensed into borosilicate vaccine vials, freeze-dried, and sealed with butyl rubber stoppers while still under reduced pressure. Vials are stored at the CDC at -20°C. The mean dry weight and SD of material in 26 randomly selected vials after freeze-drying was 22.3 ± 0.8 mg; the mean water content and its SD as determined by the Karl Fischer residual moisture test of 26 randomly selected vials were 2.67 and 0.95%, respectively, of the total weight of freeze-dried material. Thermal stability studies of the freeze-dried material indicate an extrapolated half life of 750 years when stored at -20°C.

The vial contents were sterile as determined by U.S.P. approved methods of sterility testing, negative for hepatitis-B surface antigen by radioimmunoassay, negative for antibody to HIV by Western blot; and free of rheumatoid factors as determined by latex agglutination and radioimmunoassay.

Reconstitution and Storage

Store the freeze-dried material at -20°C until use. To reconstitute, the contents should first be shaken to the bottom by tapping of the upper end. Before the stopper is removed, the vacuum should be broken by insertion of a hypodermic needle through the rubber stopper. Precisely 0.50 ml of distilled water should then be added, and the vial restoppered. The freeze-dried powder should dissolve readily with gentle swirling (avoid foam). Allow to stand for at least 1 h before use and store at 4°C until use, not later than 24 h after reconstitution. Although not recommended, the reconstituted material will withstand at least 8 weekly freeze-thaws without loss of activity. If future use of reconstituted material is contemplated, portions of the undiluted material sufficient for a single use should be stored at -70°C and discarded after use.

FANA Content

This serum gives a speckled FANA pattern due to antibodies to U1-RNP, SSB/La and SSA/Ro. Nine reference laboratories using fluorescein-labelled polyvalent (all Ig classes) α-human Ig reagents and a variety of substrates (mouse liver, mouse kidney, rat liver, and the Hep-2 human cell line) obtained a median titer of 1:320 with a modified range of 1:180 to 1:640. Using the W.H.O. fluorescein-labelled α-human immunoglobulin standard at the recommended dilution (1:20), this serum had a titer of 1:1024 using mouse liver as substrate. Using immunoglobulin class-specific reagents, titers with mouse liver as substrate were: αIgG, 1:256; αIgM, 1:16; αIgA, <1:4.

Suggested procedure for Standardization of Quality Control Reagents
The sensitivity of the FANA test is influenced by the choice of tissue substrate, the staining procedure, the fluorescein conjugate, the microscope, and the observer. Changes in any of these in the day to day performance of the test may be detected by including a stable positive control (and negative control) serum in each run.

Since the amount of AF/CDC reference preparation is limited, it should be used to calibrate secondary standards which can be run each day along with other samples being analyzed. The validity of any secondary standard depends on its having the same specificity as the primary AF/CDC standard (anti-nDNA, anti-Sm, etc.). The specificity can be determined by EIA, double immunodiffusion (anti-nRNP, anti-Sm, anti-SSB/La) or by analysis for anti-nDNA (Farr assay, millipore filter assay, Crithidia test).

To calibrate a secondary standard (i.e., a positive control to be included in subsequent routine test runs):

1. Reconstitute ampule as described above.
2. Prepare doubling dilutions of the AF-CDC reference serum and the secondary standard from 1:10 to 1:1280.
3. Carry out the routine test procedure on the dilutions.
4. Determine the nuclear fluorescence end point showing minimal recognizable staining.
5. Relative potency = \( \frac{\text{reciprocal of end point dilution of secondary standard}}{\text{reciprocal of end point dilution of AF-CDC serum}} \)

If the secondary standard is stable, the relative potency obtained should not change. Any changes in day to day results with the secondary standard are more likely due to changes in test performance.

Generally, in the day to day performance of the test, serial dilutions of the secondary standard are run. A single dilution, unless it is run at or near its end point, may not reveal major changes in test performances.

In selecting a secondary standard, a serum (available in plentiful supply) containing anti-nuclear antibody of similar specificity and resulting in a similar staining pattern should be obtained. The titer need not be similar. This should be stored undiluted in aliquots sufficient for one run (store at -20°C or below).

These reference sera can also be used to determine the most suitable or sensitive method, kit, or reagents, to compare one's results with that of the laboratories which initially evaluated the reference serum, and as a common reference for interlaboratory comparability.

Consensus evaluation results on the AF-CDC preparation are given above (FANA content) for comparison to your own results.

Caution

This serum was found to be negative for hepatitis-B surface antigen, hepatitis C and HIV antibody. Since no test method can offer complete assurance that these or other infectious agents are absent, this serum should be handled at the Biosafety Level 2 as recommended for any potentially infectious human serum or blood specimen (Centers for Disease Control, National Institutes of Health. Biosafety in Microbiological and Biomedical Laboratories, 1st Edition, 1984, 11-13).

Supplementary Information


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